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비용에서 IL-4, IL-6, GM-CSF와 IFN-γ mRNA의 발현

가 박용진·윤희로

Expression of IL-4, IL-6, GM-CSF and IFN- mRNAs in Nasal Polyps

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- ABSTRACT -

Background and Objectives : Nasal polyps are chronic inflammatory polyps originating from the middle meatus and maxillary sinus, and are usually resistant to medical treatment and often recur after surgical resections. Although the etiology and the pathogenesis are still unknown, recent studies have suggested an immunologic role for the development of nasal polyps. This study aimed to investigate the cellular infiltration and cytokine mRNA expression in nasal polyps, and compare the histopathology and cytokine profile of patients with clinically different nasal polyps. Materials and Methods : Authors examined the infiltration of lymphocytes, eosinophils, and plasma cells as well as the expression of IL-4, IL-6, IFN-, and GM-CSF mRNAs using the semiquantitative RT-PCR method in antrochoanal (n=6), allergic nasal (n=6), and nonallergic nasal polyps (n=16), and the middle turbinate of normal controls (n=4). Results : Plasma cell infiltration was more intense in nasal polyps irrespective of the type (p<0.05), and eosinophil infiltration in allergic nasal polyps was more intense than in the normal tissue (p < 0.05). Antrochoanal polyps showed higher levels of IL-4, IL-6, and GM-CSF mRNA expression, and increased IL-4/IFN- ratio, suggesting that the humoral immune reaction was predominant. Nonallergic nasal polyps showed higher levels of IFN-, and GM-CSF mRNA expression, and low IL-4/IFN- ratio, suggesting that the cellular immune reaction was predominant. Allergic nasal polyps showed higher levels of IL-4, IFN-, and GM-CSF mRNA expression, suggesting that both the cellular and humoral immune reactions may coexist. Conclusion : Different immune mechanisms may be responsible for nasal polyps of different clinical types. (J Clinical Otolaryngol 1999;10:53-60)

KEY WORDS : Nasal polyp · Cytokine · Reverse transcription polymerase chain reaction.



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대상 및 방법

des pteronyssinus and/or Dermatophagoides fari -

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RAST

cytokine mRNA

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연구대상

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radioallergosorvent test

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, IgE 7├ 450 IU/mI 4 . , , interferon - gamma(IFN -),

> interleukin(IL) - 4 - 6, granulocyte/macrophage colony stimulating factor(GM - CSF) mRNA

· 조직 염색 및 광학 현미경 관찰

, 10% para ffin hematoxylin - eosin , total RNA . , 2 7⊦ 400

. , - 3+ :(-) = 7\; /400 , (+) = 7\; 1 5 /400 , (2+) = 7\; 6 30 /400 , (3+) = 7\; 31 /400

RNA 분리 및 역전사 중합효소 연쇄반응(reverse transcription-polymerase chain reaction, RT-PCR)

RNA

 RNAzol[™] B reagent(Tel - Test, Inc.,

 Texas, U.S.A.)(2 ml/100 mg tissue)
 1.5 ml

 microcentrifuge tube
 RNA

 - 70
 .
 (Biospec pro

 ducts, Inc., Bartlesville State, U.S.A.)
 2 ml
 0.2 ml
 chl

IL - 4, IL - 6, GM - CSF IFN - mRNA

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oroform 가 15 5 15 12, 000 q mi -가 crocentrifuge tube isopropanol 4 15 4 12,000 RNA pellet 75% 15 g ethanol 10 RNA pellet diethylpyrocarbonate(DEPC) (DEPC water) GeneQuant (Pharmacia Biotech., Cambridge, England) - 70 RNA .

(Reverse transcription) total RNA cDNA Supers $cript^{TM}$ preamplification system kit(Life Technolo gies, Gaithersburg, MD, U.S.A.) . Mi crocentrifuge tube 2 µg RNA 1 µl random **DEPC** water hexamer 12 µl가 70 10 가 1 10×PCR buffer 2 µl, 25 incubation . mM MgCl₂ 2 µl, 10 mM dNTP 1 µl, 0.1 M DTT 2 25 incubation μl 10 Superscript reverse transcriptase(Life Technologies)(200 U/µI) 1 µI 25 10 , 42 water bath 50 70 15 incubation 1 가 RNase H 1 µI 37 20 incubation

Table 1. Primer sequences used for PCR

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(polymerase chain reaction, PCR) - actin, IL - 4, IL - 6, IFN - GM - CSF primer Seoul Clinical Laboratory (Seoul, Korea)

Table 1 . PCR upstream primer(100 pM) downstr eam primer(100 pM) 1 µl, cDNA 2 µl, $10 \times PCR$ buffer 5 µl, 25 mM MgCl₂ 3 µl, 10 mM dNTP 1 µl, 36.5 µl Tag DNA polymerase(Promega, Madison, WI, U.S.A.) 0.5 thermocycler(Perkin Elmer Moμl del, Branchburg, NJ, U.S.A.) . Denaturation , An-94 1 IL - 6 nealing 55 2 , IL - 4/IFN -

57 2 , GM - CSF/ - actin 66 2 , Extension 72 3 30 . cytokine cDNA

plateau . cytokine cDNA(Am erican Type Culture Collection, Maryland, U.S.A) , internal standard - actin cDNA(Amer ican Type Culture Collection, Maryland, U.S.A)

, reverse transcriptase

Cytokines	Expected size (bp)	Primer sequence		
IL-4	363	5'-GAACTITGTCCACGGACAC-3'		
		5'-GCCTTTCCAAGAAGTTTTCC-3'		
IL-6	372	5'-GAAGCTTCCAAACATGGCTGA-3'		
		5'-AGGATCCCATGCTACATTTGC-3'		
IFN-	325	5'-TICTCTIGGCTGTTACTG-3'		
		5'-ACCGAATAATTAGTCAGC-3		
GM-CSF	319	5'-CAAGCITAAGGGCCCCITGACCATG-3'		
		5'-TGGATCCGGGTCAGTGTGGCCCAGGG-3		
-actin	275	5'-GAAGCTTCCAAACATGGCTGA-3'		
		5'-AGGATCCCATGCTACATTTGC-3'		

RT - PCR (semiqu antitative analysis) (loading bu -10 µl ffer ; 0.25% bromophenol blue ; 0.25% xylene cy anol FF; 30% glycerol in water) 2 µl 0.5 µg/ml ethidium bromide7⊦ 1.5% (agarose gel) 100 V 40 Eagle Eye[™] still video . system(Stratagene, La Jolla, CA, U.S.A.) , Eagle Sight[™] 3.0 Image Capture and Anal ysis Software(Stratagene) cytokine

band intensity band intensity - actin density .

통계분석

cytokine mRNA SPSS(Windows version 7.5) software , Kruskal - Wallis , Wilkoxon rank sum test .

Chi - square test

± p<0.05

결 과

임파구, 호산구 및 형질세포의 침윤 정도 paraffin hematoxylineo sin 400 (Table 2).

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(p<0.05).

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 (p<0.05).</th>

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Table 2. Infiltration of lymphocytes, eosinophil, and pl-asma cells in subepithelial layer of nasal mucosa of no-mal control (NC), antrochoanal polyp (ACP), allergicnasal polyp (AP), and nonallergic nasal polyp (NAP)

Cell type	Grade	ACP	AP	NAP	NC
		(n=6)	(n=6)	(n = 16)	(n=4)
Lymphocyte	+	1	0	0	0
	+ +	4	3	11	4
	+ + +	1	3	5	0
Eosinophil	+	4	0	10	4
	+ +	2	2*	5	0
	+ + +	0	4*	1	0
Plasma cell	+	1	1	4	4
	+ +	5*	5*	12*	0
	+ + +	0	0	0	0

(+) = 1 - 5 cells per field ($\times 400$)

+ +) = 6 - 30 cells per field($\times 400$)

(+++) = above 30 cells per field (× 400)

*p<0.05, compared with NC.

Tissues were fixed with paraffin and stained with HE. Ce-Ils infiltrated in each section were counted at the field of 400 magnifications, and graded as described in Methods.

> 4 + . 가.

, (p<0.05) フト

가 (p<0.05) 7

Cytokine mRNA의 분석

IL-4 IFN-

RNA , RT - PCR IL - 4 IFN - . 가 cytokine 가

, IL-4 26 IFN- 28

(Fig. 1). , 가 cytokine - actin

(Fig. 2). , IL - 4 IFN -

28

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Fig. 1. Expression of IL-4 and IFN-g in nasal polyps. Total RNAs were purified from normal middle turbinates (NC, lane 1, n=4), antrochoanal polyps (ACP, lane 2, n=6), allergic nasal polyps (AP, lane 3, n=6), and nonallergic nasal polyps (NAP, lane 4, n=16), and RT-PCR was done as described in Materials and Methods. Ten microliters of each sample were separated on 1.5% agarose gel, and stained with ethidium bromide. Data shown in this figure are the representatives from each experimental group. P, positive control for PCR.



Fig. 2. Semiquantitative analysis of IL-4 and IFN-g mRNA in normal control (NC, n=4), antrochoanal polyp (ACP, n=6), Allergic nasal polyp (AP, n=6), and nonallergic nasal polyp (NAP, n=16). Ten microliters of RT-PCR products for IL-4, IFN-, and -actin were separated, and the density of each band was measured using densitometry as described in Methods. The relative amount of IL-4 and IFN- mRNA was expressed as the ratio of the density of IL-4 and IFN- to that of -actin. Data are mean \pm SD (bars). *p<0.05, compared with NCP, \pm p<0.05, compared with NAP.





Fig. 3. Semiquantitative analysis of IL-6 mRNA in normal control (NC, n=4), antrochoanal polyp (ACP, n=6), allergic nasal polyp (AP, n=6), and nonallergic nasal polyp (NAP, n=16). Ten microliters of RT-PCR products for IL-6 and b-actin were separated, and the density of each band was measured using densitometry as described in Methods. The relative amount of IL-6 mRNA was expressed as the ratio of the density of cytokine to that of -actin. Data are mean±SD (bars). *p<0.05, compared

-actin. Data are mean \pm SD (bars). "p<0.05, compared with NC, AP and NAP.



Fig. 4. Semiquantitative analysis of GM-CSF mRNA in normal control (NC, n=4), antrochonal polyp (ACP, n=6), allergic nasal polyp (AP, n=6), and nonallergic nasal polyp (NAP, n=16). Ten microliters of RT-PCR products for GM-CSF and b-actin were separated, and the density of each band was measured using densitometry as described in Methods. The relative amount of GM-CSF mRNA was expressed as the ratio of the density of cytokine to that of -actin. Data are mean \pm SD (bars). *p<0.05, compared with NC.



: IL - 4, IL - 6, GM - CSF IFN - mRNA

Table 3. Summary of the semi quantitative analysis of IL-4, IFN-g, IL-6, and GM-CSF mRNA in normal contr	ol (NC)
antrochoanal polyp (ACP), allergic nasal polyp (AP), and nonallergic nasal polyp (NAP)	

	NC	NC ACP		Middle meatal polyp		
	(n=4)	(n:	=6)	AP		NAP
IL-4	0.007	0.66 ± 0.30		0.62 ± 0.16		0.24 ± 0.27
IFN-g	0	0.10 :	± 0.05	0.42 ± 0.15		0.51 ± 0.38
IL-6	0	0.11 ± 0.12		0		0
GM-CSF	0.05 ± 0.02	0.15 ± 0.08		0.12 ± 0.02		0.15 ± 0.02
The relative amo actin. Data are n	unt of each cytokine mRNA v nean±SD.	was expresso	d as the rat	io of the density o	f each c	ytokine to that of -
			4		,	
				IFN	-	
				,		
			가	가		
IL - 4		가		IL - 4/IFN	-	
,					, IFI	N -
가	, IL-	- 4		가		Hamilos ¹⁸⁾
가가						Th - 2 cytokine
	IL - 4		IL - 6	3		가
90%가	Nonaka ¹⁷⁾			,		
		IL - 4	IL - 4	IL - 6		
				,		IL - 6
						IL - 6
				Davidssor	19) 1	
	. IL - 4 IgE			IL - 6		
가	Th - 2 cytokine IL - 5	IL -		,		IL - 6

6 IL-6 가 가가 IL - 6 • GM - CSF , IL - 5 가 GM - CSF가 가 . Hamilos 18) IL - 5 Th - 1 Th - 2 cytokine

IFN - 가 . Th - 1 cytokine IFN - Th - 2 cytokine , cytokine IL - 4 , IL -





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